Sample preparation for “Colony PCR”

- Put 20 µl of aqua dest. into each tube of a PCR tube stripe (make sure that the water is at the bottom of the tubes!)
- Pick the colonies with yellow tips and put each tip into the prepared PCR tubes, shake 10 min on a plate shaker
- Transfer the tips with a pipet to 1.5 ml Eppendorf tubes filled with 100 µl LB media (+ previously used antibiotic), pipet several times up and down and discard the tips. Shake the tubes with 400 rpm at 37°C until the results of the PCR are available. The pre-culture is later used to inoculate LB media for a Mini or Midi Prep.
- Seal the PCR stripes with caps and boil them 10 min at 95°C in a PCR cycler
- Pellet remaining bacterial material 5 min at 4000 rpm
- Use 3 µl of the supernatant as sample for a 25 µl colony PCR reaction (note: don’t disrupt pellet)